Daily sperm production, gonadal and extragonadal sperm reserves in rabbit bucks fed dietary ascorbic acid supplementation

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Target Audience: Feed producers and rabbit farmers

Abstract

A study was conducted for 20 weeks to investigate the effect of dietary ascorbic acid supplementation levels on daily sperm production, gonadal and extragonadal sperm reserves of crossbred rabbit bucks. A total of 20, matured rabbit bucks of 18 to 20 weeks old with average weight of 2.75 kg were used. Ascorbic acid was included in the diets of the rabbits at levels of 0 (control), 100, 200, 300 and 400 mg/kg feed. The rabbits were randomly allotted to the diets (n=4 rabbits/treatments) and housed individually. Each rabbit was a replicate. At the end of the experiment, two bucks per treatment were sacrificed and their testes and epididymides were carefully sampled, weighed and processed for determination of gonadal and epididymal spermatozoa reserves. All data were subjected to general linear model while the significant differences in means were separated using pairwise-difference. Results showed that gonadal sperm reserves of the right testis of bucks fed ascorbic acid supplementation were significantly (P<0.05) higher (288.75, 287.15, 299.98, 296.00 vs. 263.10 $x10^{6}$ /ml) than their counterparts fed the control diet. Bucks fed ascorbic acid supplemented diet had significantly (P<0.05) higher (98.76, 98.00, 115.72, 112.67 vs. 83.55) daily sperm production of the right testis compared to bucks fed the control diet. The results of this study suggest that feed grade ascorbic acid supplementation could improve gonadal and extragonadal sperm reserves and daily sperm production of rabbit bucks.

Key words: Ascorbic acid, rabbits, sperm reserves.

Description of problem

ambient temperature reproductive performance of rabbit bucks (1). Heat stress (due to high ambient temperature) coupled with excessive production pro-oxidants, of causes imbalance pro-oxidants between and antioxidants, resulting in oxidative stress (2). These stress are damaging agents which have been reported to induce an adaptive response in rabbits (3). (4) suggested that heat stress is induced under high ambient temperature coupled with very high relative humidity. When rabbits are exposed to heat/oxidative stress, a series of remarkable changes in their biological functions are effected which ends with reduction in productivity, libido, fertility and embryonic survival. Other effects may include facilitated disease processes from the resulting cellular damage and impairment of production and reproduction performance (5). Antioxidant

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vitamins such as ascorbic acid among others is a substance that interact with and protect cells/soft tissues against damaging effect of unstable free radical molecules (6). Ascorbic acid is essential in maintaining homeostasis such as alleviation of heat stress in rabbits. It is also essential for maturation of gonads, maintenance of tubular structure of the testes and gonadal tissue restoration (7).

This investigation was to assess the gonadal and epididymal sperm reserves and sperm production of rabbit bucks fed ascorbic acid supplementation.

Materials and methods

The study was conducted in the Rabbit Unit of the National Animal Production Research Institute (NAPRI), Shika, Zaria, situated on latitude 11° and 12°N and between longitude 7° and 8°E with an altitude of 691m above sea level (8). The mean maximum temperature varies from 19°C to 38°C depending on season, while the mean relative humidity during dry and wet season is 21% and 72% respectively. Morning temperature range between 19°C and 32°C while afternoon temperature ranges between 20°C and 40°C (9). A total of 20 sexually-mature crossbred rabbit bucks aged 18 to 20 weeks old with an average weight of 2.75 kg were used. The rabbits were weighed and randomly allotted to five dietary groups in a completely randomized design. Each group constituted a treatment and each rabbit within a group was a replicate (four rabbits per treatment). The rabbits were equally assigned to one of the following levels of ascorbic acid (0, 100, 200, 300, 400 mg ascorbic acid/kg diet).

At the end of the feeding trial, two bucks were anesthetized and sacrificed and their reproductive tracts were dissected. The testes and epididymides were carefully removed and weighed. The right testis, the left testis and the different segments of

epididymides (caput, corpus, cauda) were homogenized separately in 0.154 \M NaCl (physiological saline) at the dilution rate of 5ml/g testis. The suspensions were mixed and filtered through a double layer of sterile gauze into clean glass test tubes and the concentrations sperm therein determined by direct haemocytometric count after proper dilution (1:20 v/v) in 0.154 M NaCl (10). The concentration of sperm cells per gram of testis parenchyma, gonadal sperm reserves, daily sperm production and daily sperm production gram per parenchyma (testis) per animal were calculated as:

Concentration per gram = number of sperm cells x volume used/weight of sample Gonadal sperm reserves = concentration per gram testis x total weight of right testis $(x10^6)$

Daily sperm production = testes sperm count (gonadal sperm reserve)/time divisor. Time divisor for rabbits = 3.43

Daily sperm production/g parenchyma (testis) per animal = Gonadal sperm reserve/gross testes weight—tunica albuginea x 1/3.43 (11).

All data obtained from this study were subjected to analysis of variance (ANOVA) of (12) package and the significant differences in means were separated using pairwise-difference (P-DIFF, 12) of the same software.

Results and discussion

The effect of ascorbic acid (AA) supplementation on gonadal and epididymal sperm reserve in rabbit bucks is presented in Table 1. Gonadal sperm reserves of right testis per g testis, per testis and per paired testes as well as epididymal sperm reserves of right and left caput and caudal epididymides of bucks fed 100, 200, 300 and 400 mg/kg diets were significantly (P<0.05) higher than their counterparts fed 0 mg

AA/kg diet. Gonadal sperm reserves of left testis per g testis, per testis and paired testes/g testis as well as epididymal sperm reserves of right and left corpus epididymides were statistically (P>0.05)

similar for all levels of ascorbic acid supplementation. Higher gonadal sperm reserves were observed in bucks fed dietary ascorbic acid supplementation compared to their counterparts fed the control diet.

Table 1: Effect of ascorbic acid supplementation on gonadal and epididymal sperm reserves in rabbit bucks

	Ascorbic acid levels (mg/kg feed)						
Parameters	0	100	200	300	400	Р	
						value	
Gonadal sperm reserves	(x10 ⁶ /ml)						
Right testis per g testis	219.25±3.28 ^b	231.00±3.28a	230.17±3.28a	235.71±3.28a	233.00±3.28a	0.032	
Per testis	263.10±3.35°	288.75±3.35 ^b	287.15±3.35 ^b	299.98±3.35a	296.00±3.35a	0.040	
Left testis per g testis	218.55±8.92	231.80±8.92	233.80±8.92	234.43±8.92	232.60±8.92	0.530	
Per testis	260.07±11.92	280.48±10.92	282.89±10.92	286.00±11.92	281.43±10.92	0.199	
Paired testes per g	218.90±9.70	277.69±9.70	231.72±9.70	234.39±9.70	232.83±9.70	0.650	
testis							
Per testes	523.17±10.85b	569.23±10.85a	570.04±10.85a	585.98±10.85a	577.43±10.85a	0.018	
Epididymal sperm							
reserves (x106/ml)							
Right:							
Caput epididymides	6.70±0.39°	7.81±0.39 ^b	7.66±0.39 ^b	9.71±0.39a	8.96±0.39a	0.037	
Corpus epididymides	1.40±1.52	4.55±1.52	1.95±1.52	3.01±1.52	2.93±1.52	0.282	
Cauda epididymides	53.80±1.05°	98.92±1.05 ^b	100.00±1.05 ^b	107.54±1.05a	102.19±1.05 ^a	0.010	
Left:							
Caput epididymides	5.74±0.40°	6.60±0.40 ^b	6.63±0.40 ^b	7.55±0.40a	7.49±0.40a	0.050	
Corpus epididymides	1.14±1.80	3.48±1.80	1.63±1.80	1.49±1.80	2.19±1.80	0.243	
Cauda epididymides	50.87±0.29d	90.00±0.29°	90.60±0.29b	91.42±0.29a	90.80±0.29b	0.010	

^{abc}: Means with different superscripts in the same row are significantly (P<0.05) different

Table 2: Effect of ascorbic acid supplementation on daily sperm production and sperm production efficiency in rabbit bucks

	Ascorbic acid levels (mg/kg feed)						
Parameters	0	100	200	300	400	P value	
Daily sperm production (x10 ⁵ /ml)							
Right testis	83.55±2.14°	98.76±2.14b	98.00±2.14b	115.72±2.14a	112.67±2.14a	0.019	
Left testis	82.53±8.93	90.55±8.93	94.62±8.93	98.90±8.93	95.59±8.93	0.125	
Sperm production efficiency							
Right testis	60.88±2.50°	67.95±2.50b	67.25±2.50b	75.10±2.50a	73.90±2.50a	0.043	
Left testis	57.40±1.38 ^d	60.20±1.38c	62.99±1.38b	66.99±1.38ª	63.64±1.38 ^b	0.035	

abcd: Means with different superscripts in the same row are significantly (P<0.05) different

The results of the current study suggest that ascorbic acid possibly augmented the highly-concentrated seminal plasma ascorbic acid in the gonads/testes to protect sperm cells from being attacked by reactive oxygen species in the testes of the animals. The scavenging

potential in gonads and seminal fluid is normally maintained by adequate levels of antioxidants such as plasma vitamin C (13). Improved epididymal sperm reserves of the right and left caput and cauda epididymides observed in bucks fed ascorbic acid supplemented diet compared to bucks fed the control diet indicates that ascorbic acid enhanced the epididymide by scavenging reactive oxygen species which deteriorate and reduce sperm reserved within the epididymis. Mammalian spermatozoa are rich in highly-unsaturated fatty acids and are apparently sensitive to oxygen induced damage mediated by lipid peroxidation (14). The effects of ascorbic acid supplementation on daily sperm production and sperm production efficiency in rabbit bucks is presented in Table 2. Daily sperm production of the right testis as well as sperm production efficiency of right and left testes of bucks fed 100, 200, 300 and 400 mg/kg diets were significantly (P<0.05) higher than bucks fed 0 mg AA/kg diet. Daily sperm production of the left testis was statistically (P>0.05) similar for all levels of ascorbic acid supplementation. Higher daily sperm production was observed in the right testis of bucks fed dietary ascorbic supplementation compared to bucks fed the control diet. These results suggest that the testicular germinal epithelium was believed to be protected by ascorbic acid from oxidative stress. Degenerated testicular germinal epithelium due to oxidative stress was reported in scorbutic guinea pigs (7). Recent data suggest that ascorbic acid has defined functions in gamete biosynthesis and protection, gonadal tissue remodeling as well important component of antioxidants in seminal plasma in many species including rabbits (15). Ascorbic acid also plays vital role the prevention/reduction of the oxidation of biomolecules induced by reactive oxygen species which cause great damage to sperm cell structures (16).

Conclusion and Application

1. Based on the results obtained from this study, it is concluded that

ascorbic acid supplementation improved gonadal and extragonadal sperm reserves as well as daily sperm production and sperm production efficiency of rabbit bucks.

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References

- Liu, H., Dong, X., Tong, J. and Zhang, Q. 2011. A comparative study of growth performance and antioxidant status of rabbits when fed with or without chestnut tannins under high ambient temperature. Animal Feed Science and Technology, 164: 89-95.
- Vaibhav, P., Oberoi, P.S. and Dang, A.K. 2017. Effect of feed supplement and additives on stress mitigation in Karan Fries heifers. Veterinary World, EISSN: 2231-0916.
- 3. Dehghan, A., Arabi, M., Nahid, S. and Aminlari, M. (2010). Changes of serum reduced and oxidized glutathione in heat stressed ram. Asian Journal of Animal and Veterinary Advances, 5: 472-477.
- 4. Marai, I.F.M., El-Darawany, A.A., Fadiel, A. and Abdel-Hafez, M.A.M. (2007). Physiological traits as affected by heat stress in sheep. A Review. Small Ruminant Research, 71: 1-12.
- Sawankumar, D.R., Vasava, A.A., Pathan, M.M., Madhira, S.P., Patel, Y.G. and Pande, A.M. (2017). Effect of season on physiological, biochemical, hormonal, and oxidative stress parameters of indigenous sheep. Veterinary World, EISSN: 2231-0916. Veterinary World, EISSN: 2231-0916

- Available at www.veterinaryworld. org/Vol.10/June-2017/13.pdf.
- Hamadouche, N., Slimani, M. and Aoues, A. (2012). Beneficial effect of administration of vitamin C in amelioration of lead hepatotoxicity. Natural Science and Biology, 4 (3), 7-13.
- 7. Wang, L., Cheng, X., Li, H., Qiu, F., Yang, N. and Wang, B. 2014. Quercetin reduces oxidative stress and inhibits activation of c-Jun N-terminal kinase/activator protein-1 signaling in an experimental mouse model of abdominal aortic aneurysm. Molecular Medical Reproduction, 9:435-42.
- 8. Ovimaps, 2012. Ovi Location Map; Ovi Earth Imagery, Dated April 3, 2012.
- Iyeghe-Erakpotobor, G.T. Akinsola, O.M., Samuel, T.T. and Pewar, P.F. 2013. Growth and physiological responses of rabbit Hyla stock in a Sub-humid tropical environment. Journal of Animal Production and Research, 25: 25-36.
- 10. Egbunike GN, Holtz W, Smidt, D. 1975. Reproductive capacity of German Landrace boars. II: Sperm production rates as determined by quantitative testicular histology and from gonadal sperm reserves. Zuchthygiene, 11: 35-37.

- Amao, E.A., Jimoh, R.A. and Adeoti, T.M. 2014. Reproductive performance of rabbit does fed graded levels of Moringa oleifera leaf meal based diet. International Journal of Sciences, 3: 50-53.
- 12. SAS, 2002. SAS/STAT. User's guide: Statistics (Release 6.04 Ed). SAS Institute Inc., Cary, NC, USA.
- Guerriero, G., Trocchia, S., Abdel-Gawad, F.K. and Ciarcia, G. 2014.
 Roles of reactive oxygen species in the spermatogenesis regulation. Mini Review Article, 5(56): 1-4.
- 14. Okubanjo, O.O., Sekoni, V.O., Ajanusi, O.J., Nok, A.J. and Adeyeye, A.A. 2014. Testicular and epididymal pathology in Yankasa rams experimentally infected with Trypanosoma congolense. Asian Pacific Journal of Tropical Disease, 4(3): 185-189.
- 15. Aydin, M. and Yuksel, M. 2011. The effects of high level ascorbic acid on pregnancy rate and litter size in rats. Journal of Medicinal Plants Research, 5(13): 2695-2698.
- O'Flaherty, C. and Matsushita-Fournier, D. 2017. Reactive oxygen species and protein modifications in spermatozoa. Biology of Reproduction, 97(4): 577-585.